Diallel analysis for bodyweight involving three genotypes of Nigerian indigenous chickens

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Diallel analysis for bodyweight involving three genotypes of Nigerian indigenous chickens


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Abstract
To evaluate heterosis, reciprocal effect, general and specific combining abilities for bodyweight, a diallel crossing experiment was conducted using three genotypes of Nigerian indigenous chickens: normal (N), frizzle (F) and naked-neck (Na). A total of 601 chicks was hatched from all possible matings between the three genotypes. The chicks used in this study were hatched from a foundation stock of 90 chickens comprising 25 hens and 5 cocks for each of the three genotypes. A mating ratio of 1 male : 5 females was employed. Furthermore, data on bodyweight were scrutinized with complete diallel analysis after they had been corrected for significant effects of hatch of birds using least squares constants. The results revealed that bodyweight (BWT) was significantly influenced by genetic groups with the frizzle-naked (FNa) having the best performance at all ages (4 - 20 weeks) except at hatch, with a mean final BW T of 1173 g at 20 weeks old. The next best performing was its reciprocal, the naked-frizzle (NaF), with a mean final BWT of 1162 g. Furthermore, the FNa gave the best estimates for heterosis and specific combining ability, while the F and NNa gave the best estimates for general combining ability and reciprocal effect, respectively. Therefore, the F genotype as sire and the Na as dam provided the most suitable combination for improved BWT. Furthermore, the use of the Na genotype as dam was more suitable owing to the significant reciprocal effect.

Keywords: Combining abilities, frizzle-feathered, heterosis, naked-neck, normal-feathered

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Introduction
The poultry population in Nigeria is estimated to be 172 million, of which chickens are approximately 160 million, guinea fowl 8.3 million, ducks 1.7 million and local turkeys 1.05 million (FAOSTAT, 2011). Nigerian indigenous chickens have been described as small-bodied, slow growing, poor feed converters and poor meat animals (Ajayi, 2010). Despite these shortcomings, they are valued for their heat tolerance, meat quality, general hardiness and ability to scavenge (Petrus, 2011). However, selection in indigenous breeds has been targeted more at adaptation to tropical harsh environments and resistance to diseases than to enhanced production (Minga et al., 2004). An improvement in growth traits such as bodyweight (BWT) becomes imperative.

Genetic progress can be attained by selection and crossbreeding (Adebambo et al., 2011). Crossbreeding could lead to the production of birds that would be better in growth rate and efficiency of feed conversion without sacrificing adaptation to the indigenous environment, thereby resulting in reduced cost of production (Adebambo et al., 2011). The outcome of crossbreeding is owing to the phenomenon of heterosis, which is expressed in the performance of hybrids. Since heterosis is almost exclusively the aggregate of all single-locus dominance effects, and as these are usually beneficial, heterosis can be expected to be in a favourable direction (Kitalyi, 1999).

The test for good combining abilities is developed by generating a diallel cross, which is a set of possible combinations between several genotypes and general populations and the analysis of data from such crosses (Hayman, 1954). The general combining ability (GCA) is defined as the average performance of a line (strain, breed or genotype) in hybrid combination with other lines (Gardner & Eberhardt, 1966). The variation in GCA is owing to additive genetic variance. Specific combining ability (SCA) refers to a cross produced by a pair of lines (Adebambo et al., 2010). It indicates cases in which certain combinations
(crosses) do relatively better or worse than would be expected, based on the average performance, that is, GCA of the two lines involved in producing that combination. The variation in SCA is owing to non-additive genetic variance, heterosis, dominance, over-dominance and epistasis (Singh & Kumar, 1994).

Large variations have been reported among the birds in morphological, immune response, growth and reproductive traits (Peters, 2000; Msoffe et al., 2001). This has led to the conclusion that indigenous chickens are repositories of unique genes that could be used in other parts of the world (Adebambo, 2005), hence the need for their conservation to keep genetic variation in and between indigenous breeds. A series of reports on the characterization of indigenous chickens revealed that they could be classified based on the occurrence of some major genes, such as the dwarf gene, naked-neck gene and frizzling gene (Ikeobi et al., 1996). Naked-neck and frizzle birds have been found to be thermally stress tolerant compared with their normally feathered counterparts (Nwachukwu et al., 2006). The naked-neck and frizzle genes have been found to be associated with heat tolerance, and therefore in areas with high ambient temperature, birds with these genes are superior to their normally feathered counterparts for feed efficiency (Garces et al., 2001). According to Fayeye et al. (2006), birds with the naked-neck and frizzle genes have better adult bodyweights than their normally feathered counterparts. Furthermore, many investigators have confirmed the superiority of crossbreds over purebreds regarding growth traits in chicken (Khalil et al., 2004; Nwachukwu et al., 2006).

There is a dearth of information on diallel analysis for BWT involving normal-feathered, frizzle-feathered and naked-neck Nigerian indigenous chickens. This study was therefore designed to investigate the heterosis, GCA, SCA and reciprocal effects for bodyweight of three genotypes of Nigerian indigenous chicken in a diallel crossing experiment.

Materials and Methods

The study was conducted at the Experimental and Research Farm of the Department of Animal Science, Faculty of Agriculture, Ahmadu Bello University, Zaria, Nigeria. The area is situated between latitudes 11°N and 12°N and an altitude of 640 m (Akpa et al., 2013).

A total of 90 sexually matured indigenous chickens, comprising 25 hens and five cocks, each of the three genotypes (normal-feathered, frizzle-feathered and naked-neck) was used as the foundation stock for this study. This foundation stock was randomly purchased from Katsina (Funtua), Kaduna (Shika, Giwa) and Kogi (Anyigba, Okene) States. The birds were fed breeder mash (10.54 MJ ME/kg and 180 g crude protein/kg), and water was offered ad libitum. They were raised on deep litter and pen mating was carried out using a mating ratio of 1 male : 5 females.

Fertile eggs were collected, stored for a maximum of seven days, sorted out sire-wise, marked according to sire identification numbers and within genotypes. The eggs were fumigated with potassium permanganate and hydrogen peroxide in the fumigation room for about 30 minutes and later taken to the incubator at a temperature of about 37 °C. Candling the eggs was done on the 18th day of incubation, and fertile eggs were transferred to the hatcher. Hatching trays were partitioned into compartments according to sire numbers and genotypes. Eggs belonging to the same sire were put in the same compartment. Hatching was completed on the 21st day of incubation and resulted in the F1 generation.

A total of 601 chicks was hatched from all possible matings among the three genotypes. The possible matings are explained in Table1. The chicks were wing-tagged and pedigreed according to sire identification number, and brooded with the aid of electric bulbs. They were fed chick mash (11.9 MJ ME/kg and 210 crude protein/kg) from 0 to 8 weeks old and grower mash (10.04 MJ ME/kg and 160 g crude protein/kg) from eight weeks old until the end of the study (20 weeks old). They were also vaccinated against common poultry diseases such as Newcastle, infectious bursal (Gumboro) and fowl-pox. Other routine medication and management operations included anti-coccidial medication, de-worming and delousing.

Bodyweight (g) was measured with a sensitive (1 g) top-loading scale (Citizen Electronic Balance, Goldair®). Bodyweight was recorded at day old and at four-week intervals until 20 weeks old. The general linear model procedure of Statistical Analysis System program (SAS, 2002) was used to test the effects of the genetic groups and sex. Significant means were separated using Tukey's studentized range (HSD) test. The following model was used to analyse the BWT trait:

\[ Y_{ijk} = \mu + G_i + S_j + e_{ijk} \]

where:  
\( Y_{ijk} \) = bodyweight of bird of the jth sex in the ith genetic group  
\( \mu \) = overall mean  
\( G_i \) = effect of the ith genetic group
Sj = effect of the jth sex
\( e_{ijk} \) = random error.

**Table 1** Mating scheme of foundation stock for the production of F1 generation

<table>
<thead>
<tr>
<th>Male</th>
<th>Female</th>
<th>Progeny</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pure cross</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Normal feathered</td>
<td>x</td>
<td>Normal</td>
</tr>
<tr>
<td>Frizzle feathered</td>
<td>x</td>
<td>Frizzle</td>
</tr>
<tr>
<td>Naked-neck</td>
<td>x</td>
<td>Naked</td>
</tr>
<tr>
<td>Main cross</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Normal feathered</td>
<td>x</td>
<td>Frizzle</td>
</tr>
<tr>
<td>Frizzle feathered</td>
<td>x</td>
<td>Naked</td>
</tr>
<tr>
<td>Naked-neck</td>
<td>x</td>
<td>Normal</td>
</tr>
<tr>
<td>Reciprocal cross</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Frizzle feathered</td>
<td>x</td>
<td>Normal</td>
</tr>
<tr>
<td>Naked-neck</td>
<td>x</td>
<td>Frizzle</td>
</tr>
<tr>
<td>Normal feathered</td>
<td>x</td>
<td>Naked</td>
</tr>
</tbody>
</table>

Heterosis was calculated according to Fairfull (1990) by applying the following formula:

\[ H\% = \frac{AB - (0.5AA+0.5BB)}{(0.5AA+0.5BB)} \times 100. \]

Where:
- \( H\% \) = percent heterosis
- \( AB \) = crossbred
- AA and BB = sire and dam genotypes, respectively.

The diallele 1 procedure of plant breeding package of R Core Team (2013) was used to estimate the general combining ability (GCA), specific combining ability (SCA) and reciprocal effects (RE) according to Griffing’s (1956) method I, using a random model (Model II). The following model was used:

\[ Y_{ijk} = \mu + g_i + g_j + s_{ij} + r_{ij} + e_{ijk} \]

where:
- \( Y_{ijk} \) = observed value of each experimental unit
- \( \mu \) = population mean
- \( g_i \) = GCA effect for \( i^{th} \) genotype
- \( g_j \) = GCA effect for \( j^{th} \) genotype
- \( s_{ij} \) = SCA effect for the cross \( (i, j) \)
- \( r_{ij} \) = RE effect of the cross \( (i, j) \)
- \( e_{ijk} \) = random observation error among replicates.

**Note:** Preliminary analysis was carried out to test for effect of hatch. Where it was found to have a significant effect, the data were transformed by adding or subtracting least square constants according to Harvey’s (1966) method.

**Results**

Table 2 shows the least square means and coefficient of variation for BWT of indigenous chickens by age across genetic groups. At day old, frizzled (F) recorded the BWT of 25.8 g, which was higher \( (P < 0.01) \) than that of normal-naked (NNa) (22.6 g). Bodyweight at 4 weeks old was lower \( (P < 0.01) \) in naked-normal (NaN), naked-neck (Na), normal (N) and naked-frizzle (NF) than in the frizzle (F) and frizzled-naked (FNa) who recorded the BWTs. At 8 weeks old, F recorded BWT of 345.2 g which was higher \( (P < 0.05) \) than NaN (305.6 g) and NNa (310.6 g) which recorded the lowest BWT. FNa recorded highest \( (P < 0.01) \) BWTs at 12,
16 and 20 weeks old while NaN was consistently the lowest. The coefficient of variation (%) ranged from 14.92 (BWT0) to 22.01 (BWT4).

Heterotic estimates for BWT from day old to 20 weeks old are presented in Figure 1. Results revealed that heterosis was negative only from day old to 8 weeks old in NF and NaF while it was negative in FN up to 12 weeks old. Heterosis was positive at all ages in FNa, except at day old, while it was negative at all ages in NaN except at day old. In NNa, heterosis was positive at all ages except at day old and 8 weeks old.

![Figure 1](image)

**Figure 1** Heterotic estimates for bodyweight from day old to 20 weeks old.

The crossbreeding genetic estimates for GCA, SCA and RE for BWT at day old to 20 weeks old are presented in Table 3. The crossbreeding genetic estimates obtained for GCA were positive at 4 and 8 weeks old only. However, the values were significant ($P < 0.05$) only at 4 weeks old. Similarly, the crossbreeding
Table 2 Least square means and coefficient of variation for bodyweight (g) from 0 to 20 weeks old across genetic groups of Nigerian indigenous chickens

<table>
<thead>
<tr>
<th>Traits</th>
<th>NO (601)</th>
<th>BWT0</th>
<th>BWT4</th>
<th>BWT8</th>
<th>BWT12</th>
<th>BWT16</th>
<th>BWT20</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Purebred</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N</td>
<td>101</td>
<td>25.5ab ± 0.36</td>
<td>101.5b ± 2.22</td>
<td>324.8abc ± 6.88</td>
<td>584.5bc ± 12.40</td>
<td>723.6d ± 15.95</td>
<td>1083.0abcd ± 22.74</td>
</tr>
<tr>
<td>F</td>
<td>55</td>
<td>25.8a ± 0.49</td>
<td>111.5b ± 3.12</td>
<td>345.2ab ± 9.95</td>
<td>584.9bc ± 17.94</td>
<td>741.5cd ± 23.07</td>
<td>1062.2bcd ± 32.88</td>
</tr>
<tr>
<td>Na</td>
<td>73</td>
<td>24.5bc ± 0.43</td>
<td>100.6c ± 2.53</td>
<td>313.7bc ± 7.89</td>
<td>577.9bc ± 12.23</td>
<td>763.0abcd ± 18.29</td>
<td>1071.9abcd ± 26.08</td>
</tr>
<tr>
<td><strong>Main crosses</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NF</td>
<td>38</td>
<td>23.9c ± 0.59</td>
<td>103.1b ± 3.88</td>
<td>315.7bc ± 11.66</td>
<td>611.0ab ± 21.04</td>
<td>799.2abc ± 27.06</td>
<td>1157.3abcd ± 38.56</td>
</tr>
<tr>
<td>FNa</td>
<td>74</td>
<td>23.5cd ± 0.43</td>
<td>112.1a ± 2.74</td>
<td>340.2ab ± 8.39</td>
<td>646.3a ± 15.13</td>
<td>831.5a ± 19.45</td>
<td>1173.2a ± 27.73</td>
</tr>
<tr>
<td>NaN</td>
<td>82</td>
<td>25.3abc ± 0.40</td>
<td>98.4b ± 3.88</td>
<td>305.6c ± 8.10</td>
<td>550.0c ± 14.57</td>
<td>716.3d ± 18.74</td>
<td>1029.7d ± 26.71</td>
</tr>
<tr>
<td><strong>Reciprocal crosses</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FN</td>
<td>52</td>
<td>25.2ab ± 0.51</td>
<td>104.7ab ± 3.34</td>
<td>316.0bc ± 9.73</td>
<td>568.9a ± 17.56</td>
<td>758.4abd ± 22.58</td>
<td>1093.3abcd ± 32.18</td>
</tr>
<tr>
<td>NaF</td>
<td>72</td>
<td>23.7cd ± 0.43</td>
<td>104.9ab ± 2.85</td>
<td>316.9a ± 8.59</td>
<td>597.7abc ± 15.50</td>
<td>817.4bc ± 19.93</td>
<td>1161.9abc ± 28.41</td>
</tr>
<tr>
<td>NaN</td>
<td>54</td>
<td>22.6d ± 0.50</td>
<td>103.6a ± 3.28</td>
<td>310.6a ± 10.07</td>
<td>616.1bc ± 18.16</td>
<td>792.3abc ± 23.35</td>
<td>1089.1abcd ± 33.28</td>
</tr>
<tr>
<td>SEM</td>
<td></td>
<td>0.15</td>
<td>0.95</td>
<td>2.92</td>
<td>5.23</td>
<td>6.70</td>
<td>9.57</td>
</tr>
<tr>
<td>CV %</td>
<td></td>
<td>14.92</td>
<td>22.01</td>
<td>20.62</td>
<td>20.10</td>
<td>19.83</td>
<td>19.77</td>
</tr>
<tr>
<td>LOS</td>
<td></td>
<td>**</td>
<td>**</td>
<td>*</td>
<td>**</td>
<td>**</td>
<td>**</td>
</tr>
</tbody>
</table>

abcd Means with different superscripts on the same column are significantly different; BWT0, BWT4, BWT8, BWT12, BWT16 and BWT20 mean bodyweights at 0, 4, 8, 12, 16 and 20 weeks, respectively.


SEM: standard error of means; CV: coefficient of variation; LOS: level of significance.

* P <0.05; ** P <0.01; NO: number of observations.
genetic estimates for RE were negative at 8 weeks old only, but the values were significant \((P<0.05)\) at day old and 12 weeks old only. The crossbreeding genetic estimates for SCA for BWT were negative only at the 4 weeks old. However, the values were significant \((P<0.05)\) at 16 weeks old and \((P<0.01)\) at 20 weeks old.

Table 3 Crossbreeding genetic estimates for general, specific combining abilities and reciprocal effects for bodyweight at day old to 20 weeks old

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Day old</th>
<th>Week 4</th>
<th>Week 8</th>
<th>Week 12</th>
<th>Week 16</th>
<th>Week 20</th>
</tr>
</thead>
<tbody>
<tr>
<td>GCA</td>
<td>−0.03</td>
<td>8.88*</td>
<td>36.4</td>
<td>−169.4</td>
<td>−23.5</td>
<td>−703.5</td>
</tr>
<tr>
<td>SCA</td>
<td>0.36</td>
<td>−4.48</td>
<td>26.2</td>
<td>477.1</td>
<td>873.1*</td>
<td>4040.0**</td>
</tr>
<tr>
<td>RE</td>
<td>0.48*</td>
<td>0.21</td>
<td>−5.9</td>
<td>456.5*</td>
<td>299.9</td>
<td>232.6</td>
</tr>
</tbody>
</table>

GCA: general combining ability; SCA: specific combining ability; RE: reciprocal effect.

* \(P<0.05\); ** \(P<0.01\).

Figure 2 depicts the estimates of GCA, SCA and RE for BWT at various ages. GCA estimates for N were negative at all ages except for day old, while F was negative only at the 12th week. Na showed negative estimates from hatch to the 8th week, but were positive from the 12th week to the 20th week. SCA was negative for NF, except at the 12th week, positive for NaN only at the 12th and 16th week but positive through all periods and increased progressively from hatch to 20 weeks for FNa. RE estimates were positive from hatch to the 8th week only for FN, negative only at hatch for NNa, while for NaF positive only at day old, 4th and 20th week.

Discussions

The range of day-old BWT in this study was within that of 22.3 g to 30.9 g reported by Oke (2011) in three genotypes of Nigerian indigenous chickens and their crosses. It was also similar to the range of 21.8 to 28.1 g recorded by Ndofor-Foleng et al. (2010) for light and heavy ecotype chickens with their main crosses. The significantly higher day-old bodyweight in F suggests that the gene may have the potential for rapid growth. This is in accordance to the report of Oke (2011), who also reported ascendant day-old weight in F and FN.

The range for BWT4 in this study fell slightly below the 101.4 - 129.2 g recorded by Oke (2011) and was higher than the 86.3 - 91.9 g reported by Egahi et al. (2013). This can be attributed to the differences in experimental setup and environment. The findings in the present study, where there was significant variation in BWT4 among genetic groups, contradicted Mahrous (2008) and Oke (2011), who had previously reported that there are no differences between the main and reciprocal crosses for bodyweight at this age. The FNa and F genetic groups, which had the highest weights, were consistent with the report of Mahrous et al. (2008) that the duo of F and Na genes conferred fast-growing ability, either singly or in pairs. Correspondingly, F and FNa genetic groups retained far superior performance of BWT8, indicating excellent individual and combining patterns for these major genes. BWT12 to BWT20 showed that the FNa and its reciprocal NaF out-performed all other groups. These observations agree with the findings of Mahrous (2008) and Oke (2011). This ascendancy may be as a result of efficiency of thermoregulation imparted by the two genes (Yunus & Cahaner, 1999).

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Range of values obtained for these periods compared favourably with the findings of Egahi et al. (2013), who reported values of 500.5 - 639.5 g, 734.4 - 842.3 g, and 1017.6 - 1121.8 g for BWT12, BWT16 and BWT20, respectively. The coefficient of variation for BWT by period was highest at BWT4 indicating that BWT varied more at 4 weeks old among genetic groups. The superior performance of F and Na with their crosses over the N throughout the study for this trait agrees with Galal (2000) that there exists superiority of naked-neck and frizzled genes over normal-feathered chickens in BWT, breast girth and tibia length.
The range of values (−9.44% to 13.48%) obtained for heterosis in BWT is similar to the findings of Iraqi et al. (2011) who reported a range of −5.24% to 9.05% when crossing occurred between Matrouh and
Inshas chicken breeds, although results obtained in the present study yielded higher heterotic percentages than those reported by the authors. Positive heterosis is an indication that the crossbred exceeded the range of variation in BWT exhibited by its parents, while negative heterosis indicates that the crossbred fell below the range of parental variation. Reports of Wilham & Pollak (1985) indicated that the magnitude of heterosis is inversely related to the degree of genetic resemblance between parental populations and is expected to be proportional to the degree of heterozygosity of the crosses (Sheridan, 1981). Thus, heterosis is a result of non-additive genetic effects and may be viewed as overall fitness, as well as an expression of a specific trait. According to Saadey et al. (2008), the presence of positive and high heterotic estimates at most ages may be an encouraging factor for breeders to use such crosses for the trait under consideration. This therefore, indicates that a cross between F males and Na females may be most suitable for BWT improvement in Nigeria since FNa had positive and high heterotic estimates for most ages.

The importance of additive genes in expression of BWT was observed by the significant GCA effect observed at 4th week of age which is indicative of the importance of additive genes for the expression of BWT trait (Afifi et al., 2002). The superior performance of the frizzle birds suggest that it has the upmost preponderance of genes which impact additive gene effect on BWT. Thus, the frizzle gene could lead to an increase in BWT because of the higher GCA values. Furthermore, Na showed positive GCA from week 12 to 20. Observed positive estimates for F and Na for most of the periods compared with N, which was negative for all periods except at hatch, were partly consistent with previous reports (Ndeyebo, 2011) that the Na showed positive GCA estimates and indicated that the F and Na possess higher gene variation for BWT than the N chicken (Adebambo et al., 2010).

The estimates obtained for SCA were significant at the 16th and 20th weeks of age, which is an indication of non-additive gene action (Kabir et al., 2012). Thus, such traits could be improved by utilising non-additive gene effects such as dominance and epistatic/maternal effects and improved management practises. Furthermore, the positive SCA values for only FNa combinations throughout the periods may indicate the suitability of these two genotypes in combination for genetic improvement and gain.

The significant estimates obtained for reciprocal effect at hatch and at the 12th week of age imply that there was variation owing to sex-linked genes and maternal effect. The superior performance of NNA may be as a result of maternal effect of Na. This result was consistent with the positive estimates obtained between Venda (normal-feathered South Africa indigenous chicken) and naked-necks (Ndeyebo, 2011). However, Cook et al. (1972) suggested that differences among male progeny of reciprocal crosses may be attributable to maternal effects and not to sex-linkage, because the homogametic males in reciprocal crosses have comparable sex chromosomes. Hence, Cook et al. (1972) stated that reciprocals test the possibility that sex-linkage is operative when significant differences are found among female progeny because each female receives its sex chromosome from its sire. The magnitude of sex linkage effects is expected to be influenced by the breeds implicated in the crossbreeding scheme, which confirms the importance of the choice breed of sire and the breed of dam in planning crossing programmes (Sabri et al., 2000).

Conclusions

The diallel analysis of three genotypes of Nigerian indigenous chickens showed that BWT was affected by genetic group. The FNa group had superior BWT compared with other genetic groups at most ages. Furthermore, the FNa gave the best estimates for heterosis and SCA, while F and NNA gave the best estimates for GCA and RE, respectively. Therefore, the use of the F as sire and the Na as dam provided the most suitable combination for the best improved BWT with a high BWT at 20 weeks old. The use of Na as dam was also most suitable since sex-linked/maternal effect was significant. In view of the estimates for GCA, SCA and RE in this study, the combination of the frizzling and naked genes as sires and dams, respectively, is highly recommended to bring about genetic improvement for BWT.

References


